

Effects of vaccination against foot-and-mouth disease virus on reproductive performance of *Bos indicus* beef cows

L. C. L. Ferreira,* R. F. Cooke,^{†‡}¹ R. S. Marques,[†]
H. J. Fernandes,[§] C. E. Fernandes,[#] R. Stelato,^{||} G. L. Franco,* and R. A. A. Lemos*

*Faculdade de Medicina Veterinária e Zootecnia, Universidade Federal de Mato Grosso do Sul, Campo Grande 79074-460, Brazil; [†]Eastern Oregon Agricultural Research Center, Oregon State University, Burns 97720; [‡]Programa de Pós-Graduação em Zootecnia, Faculdade de Medicina Veterinária e Zootecnia, Universidade Estadual Paulista, Botucatu 18618-970, Brazil; [§]Unidade Universitária de Aquidauana, Universidade Estadual de Mato Grosso do Sul, Aquidauana 79200-000, Brazil; [#]Centro de Ciências Biológicas e da Saúde, Universidade Federal de Mato Grosso do Sul, Campo Grande 79074-460, Brazil; and ^{||}Laboratório Zoetis Ltda., Campo Grande 79074-460, Brazil

ABSTRACT: This study compared reproductive performance of *Bos indicus* cows vaccinated against the foot-and-mouth disease (FMD) virus before timed AI or during early pregnancy (Exp. 1), as well as rectal temperature (RT) and plasma concentrations of the acute-phase protein haptoglobin in cattle vaccinated or not against the FMD virus (Exp. 2). Cattle utilized in Exp. 1 and 2 originated from herds with no historical occurrences of FMD and that received vaccination against the FMD virus biannually. In Exp. 1, 604 lactating, multiparous, nonpregnant Nelore cows were randomly assigned on d -31 of the experiment to receive 1) vaccination against the FMD virus on d \geq 31 (VACPRE; $n = 291$) and 2) vaccination against FMD virus on d 30 (VACGEST; $n = 313$). From d -11 to 0, all cows were assigned to an estrus synchronization + timed AI (d 0) protocol. Pregnancy status to AI was verified on d 30 and 90 via transrectal ultrasonography. A treatment \times day interaction was detected ($P < 0.01$) for pregnancy rates to AI, which were similar ($P = 0.17$) between VACPRE and VACGEST on d 30 (61.8% vs. 56.2%, respectively; SEM = 2.8) but greater ($P < 0.01$) for VACPRE on d 90 (59.4% vs. 46.9%, respectively; SEM = 2.8). Pregnancy loss from d 30 to 90 was greater ($P < 0.01$) in

VACGEST compared with VACPRE (16.5% vs. 3.9%, respectively; SEM = 2.2). In Exp. 2, 40 pregnant Nelore females (20 nulliparous and 20 multiparous cows; BCS = 4.73 ± 0.12) were ranked by parity and assigned to receive (VAC; $n = 20$) or not receive (NOVAC; $n = 20$) vaccination against the FMD virus. Blood samples were collected and RT was recorded before (h 0) and 24, 72, 120, and 168 h after treatment administration. Treatment \times day interactions were detected ($P < 0.01$) for RT and plasma haptoglobin. The RT was greater ($P < 0.01$) in VAC compared with NOVAC at 24 h after treatment administration and was similar ($P \geq 0.31$) between treatments at all other sampling hours. Plasma haptoglobin concentration was similar ($P = 0.98$) between VAC and NOVAC before treatment administration ($P = 0.48$) and greater ($P < 0.01$) in VAC at 24, 72, 120, and 168 h after treatment administration. In summary, vaccinating *B. indicus* beef cows against the FMD virus resulted in a 4-fold increase in pregnancy loss when the vaccine was administered 30 d after timed AI compared with 31 d before timed AI. These outcomes can be associated with inflammatory and acute-phase reactions elicited by the FMD vaccine, which are known to impair pregnancy maintenance in cattle.

Key Words: acute-phase response, beef cattle, foot-and-mouth disease, inflammation, pregnancy loss

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INTRODUCTION

Foot-and-mouth disease (FMD) is a severe, highly contagious viral disease that affects cloven-hoofed livestock species, including cattle (Grubman and Baxt, 2004), and has been recognized as a major constraint to international trade in animals and animal products

¹Corresponding author: reinaldo.cooke@oregonstate.edu
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(Leforban, 1999). Although FMD has been eradicated in North America and western Europe, this disease is still endemic in Africa, South America, and Asia (Animal and Plant Health Inspection Service, 2013). Vaccination against the FMD virus successfully reduced FMD outbreaks in many parts of the world (Brown, 1992; Kahn et al., 2002); therefore, vaccination is a common and often mandatory strategy used to mitigate FMD in endemic regions (Rodriguez and Gay, 2011).

Early pregnancy loss, particularly during the first trimester of gestation, is a major reproductive challenge in cow-calf systems (Humblot, 2001). Hence, strategies to alleviate early pregnancy losses are warranted for optimal reproductive and overall efficiency of cow-calf operations. The majority of FMD vaccines used worldwide contain inactivated FMD virus serotypes and an oil-based adjuvant to elicit greater immune protection to target antigens (Rodriguez and Grubman, 2009). In general, adjuvants elicit innate immune responses associated with antigen presentation to T-cell lymphocytes, including inflammatory and acute-phase reactions (Tizard, 2004; Rodrigues et al., 2015) known to result in pregnancy losses in cattle (Hansen et al., 2004). Using this rationale, we hypothesized that administration of a FMD vaccine during early pregnancy stimulates an acute-phase protein reaction and results in increased pregnancy loss in vaccinated cattle. To test this hypothesis, Exp. 1 compared reproductive performance of *Bos indicus* cows vaccinated against the FMD virus before timed AI or during early pregnancy, whereas Exp. 2 compared rectal temperature and plasma concentrations of the acute-phase protein haptoglobin in cattle vaccinated or not vaccinated against the FMD virus.

MATERIALS AND METHODS

Experiment 1 was conducted on a commercial cow-calf operation located in Miranda, Brazil, and cattle were cared for in accordance with the practices outlined in the Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching (Federation of Animal Science Societies, 2010). Experiment 2 was conducted at the Universidade Federal do Mato Grosso do Sul, located in Terenos, Brazil, and cattle were cared for in accordance with acceptable practices and experimental protocols reviewed and approved by the Universidade Federal do Mato Grosso do Sul Animal Ethics Committee. Cattle utilized in Exp. 1 and 2 originated from herds with no historical occurrences of FMD that received vaccination against the FMD virus twice a year. In addition, all cattle utilized herein were vaccinated against FMD approximately 6 mo before the beginning of each experiment.

Experiment 1

Animals and Treatments. A total of 604 lactating, multiparous, nonpregnant Nelore cows (approximately 65 to 95 d days postpartum; BCS = 3.85 ± 0.05 according to Wagner et al., 1988), maintained in 2 groups of 266 and 338 cows each, were assigned to the experiment (d -31 to d 90 relative to timed AI). Groups were maintained in individual *Brachiaria brizantha* pastures with ad libitum access to water and a commercial mineral-vitamin mix (DSM Produtos Nutricionais Brasil, São Paulo, Brazil) and were independently assigned to experimental procedures 1 d apart. Within each group, cows were randomly assigned on d -31 of the experiment to receive 1) vaccination against the FMD virus (5 mL subcutaneous [s.c.] of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, Brazil) on d -31 of the experiment (**VACPRE**; 31 d before timed AI) or 2) vaccination against the FMD virus (5 mL s.c. of Ourovac Aftosa; Ourofino Saúde Animal) on d 30 of the experiment (**VACGEST**; 30 d after timed AI).

On d -11, both groups were assigned to the same estrus synchronization plus timed AI protocol (Meneghetti et al., 2009; d -11 to 0). More specifically, cows received a 2-mg injection (intramuscular [i.m.]) of estradiol benzoate (Gonadiol; Zoetis, São Paulo, Brazil) and an intravaginal progesterone-releasing device (**CIDR**, containing 1.9 g of progesterone; Zoetis) on d -11. On d -2, CIDR was removed, and cows received a 12.5-mg injection (i.m.) of PGF_{2 α} (Lutalyse; Zoetis) in addition to a 0.6-mg injection (i.m.) of estradiol cypionate (Zoetis) and 300 IU injection (i.m.) of eCG (Novormon; Zoetis). On d 0, cows were assigned to timed AI. All cows were inseminated with semen from a single sire. Within each group, cows were inseminated by 1 of 2 technicians, and the distribution of cows inseminated by each technician was equal within each treatment.

Sampling. Cow BCS (Wagner et al., 1988) was assessed at timed AI on d 0. Pregnancy status to AI was verified on d 30 and 90 of the experiment by detecting a viable conceptus with transrectal ultrasonography (5.0-MHz transducer; Chison 600, Chison Medical Imaging Co., Ltd., Wuxi, China). Cows were not exposed to bulls or to additional AI services between timed AI and pregnancy evaluation on d 30. On d 31, cows were exposed to mature bulls for 50 d (1:25 bull to cow ratio). Cows diagnosed as pregnant on d 30 and then nonpregnant or with an estimated conceptus age of ≤ 60 d on d 90 (Curran et al., 1986) were considered to have lost the AI pregnancy.

Statistical Analysis. Quantitative and binary data were analyzed, respectively, with the MIXED and GLIMMIX procedures of SAS (SAS Inst. Inc., Cary, NC; version 9.3) and the Satterthwaite approximation to determine the denominator degrees of freedom for the tests of fixed effects, using cow as the experimen-

tal unit and cow(treatment \times group) as the random variable. The model statement used for analysis of cow BCS on d 0 and pregnancy loss contained the effects of treatment, group, and the resultant interaction. The model statement used for analysis of pregnancy rates to timed AI contained the effects of treatment, group, day of pregnancy diagnosis (d 30 or 90), and all resultant interactions. Results are reported as least squares means and separated using LSD. Significance was set at $P \leq 0.05$, and tendencies were determined if $P > 0.05$ and ≤ 0.10 . Results are reported according to treatment effects if no interactions were significant or according to the highest-order interaction detected.

Experiment 2

Animals and Treatments. A total of 40 pregnant Nelore females, including 20 nonlactating nulliparous and 20 lactating multiparous cows, were assigned to the experiment (BCS = 4.73 ± 0.12 according to Wagner et al., 1988). All cows were maintained in a single *B. brizantha* pasture with ad libitum access to water and a commercial mineral-vitamin mix (DSM Produtos Nutricionais Brasil). At the beginning of the experiment (d 0), cows were ranked by parity and assigned to receive 1) vaccination against the FMD virus (VAC; 5 mL s.c. of Ourovac Aftosa; Ourofino Saúde Animal) on d 0 or 2) no vaccination against the FMD virus on d 0 (NOVAC).

Sampling. Rectal temperature was recorded (G-Tech digital thermometer; G-Tech, São Paulo, Brazil) and blood samples were collected immediately before (h 0) and 24, 72, 120, and 168 h after treatment administration. Blood was collected via jugular venipuncture into commercial blood collection tubes (Vacutainer, 10 mL; Becton Dickinson, Franklin Lakes, NJ) with 158 USP units of freeze-dried sodium heparin, placed immediately on ice, centrifuged ($2,500 \times g$ for 30 min; 4°C) for plasma harvest, and stored at -20°C on the same day of collection. All plasma samples were analyzed for haptoglobin concentration according to colorimetric procedures described by Cooke and Arthington (2013). The intra- and interassay CV were, respectively, 2.4% and 7.6%.

Statistical Analysis. Data were analyzed with the MIXED procedure of SAS (SAS Inst. Inc.; version 9.3) and the Satterthwaite approximation to determine the denominator degrees of freedom for the tests of fixed effects, using cow as the experimental unit and cow(treatment \times parity) as the random variable. The model statement used for analysis of plasma haptoglobin and rectal temperature contained the effects of treatment, parity, hour, and all resultant interactions. The specified term for the repeated statements was hour, cow(treatment \times parity) was the subject, and the covariance structure utilized was autoregressive on the basis of the Akaike infor-

Table 1. Reproductive performance of *Bos indicus* beef cows vaccinated against the foot-and-mouth disease virus (5 mL subcutaneous of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, Brazil) on d -31 (VACPRE; $n = 291$) or d 30 (VACGEST; $n = 313$) relative to timed AI (d 0)¹

Item	VACPRE	VACGEST	SEM	P-value
Pregnancy rates to timed AI, ² %				
d 30	61.8 (180/291)	56.2 (176/313)	2.8	0.17
d 90	59.4 (173/291)	46.9 (147/313)	2.8	<0.01
Pregnancy loss from d 30 to 90, ³ %	3.9 (7/180)	16.5 (29/176)	2.2	<0.01

¹On d -11, groups were assigned to the following estrus synchronization plus timed AI protocol. Cows received a 2-mg injection (intramuscular [i.m.]) of estradiol benzoate (Gonadiol; Zoetis, São Paulo, Brazil) and an intravaginal progesterone-releasing device (CIDR, containing 1.9 g of progesterone; Zoetis) on d -11. On d -2, CIDR was removed, cows received a 12.5-mg injection (i.m.) of PGF_{2 α} (Lutalyse; Zoetis) in addition to a 0.6-mg injection (i.m.) of estradiol cypionate (Zoetis) and a 300-IU injection (i.m.) of eCG (Novormon; Zoetis). On d 0, cows were assigned to timed AI. Pregnancy status to AI was verified on 30 and 90 d after timed AI by detecting a viable conceptus with transrectal ultrasonography (5.0-MHz transducer; Chison 600, Chison Medical Imaging Co., Ltd., Wuxi, China).

²Values within parentheses represent the number of pregnant cows divided by the number of total cows assigned to timed AI.

³Values within parentheses represent the number of cows that lost AI pregnancy divided by the number of diagnosed as pregnant to timed AI on d 30.

mation criterion. Results are reported as least squares means and separated using LSD. Significance was set at $P \leq 0.05$, and tendencies were determined if $P > 0.05$ and ≤ 0.10 . Results are reported according to treatment effects if no interactions were significant or according to the highest-order interaction detected.

RESULTS

Experiment 1

No treatment differences were detected ($P = 0.87$) for cow BCS at timed AI (3.87 vs. 3.84 BCS for VACPRE vs. VACGEST cows; SEM = 0.10).

A treatment \times day interaction was detected ($P < 0.01$) for pregnancy rates to AI (Table 1), which were similar ($P = 0.17$) between treatments on d 30 but greater ($P < 0.01$) for VACPRE compared with VACGEST cows on d 90. Accordingly, pregnancy loss from d 30 to 90 was greater ($P < 0.01$) in VACGEST compared with VACPRE cows (Table 1).

Experiment 2

A treatment \times hour interaction was detected ($P < 0.01$) for rectal temperature, which was similar between VAC and NOVAC cows before treatment administration ($P = 0.48$), greater ($P < 0.01$) in VAC compared

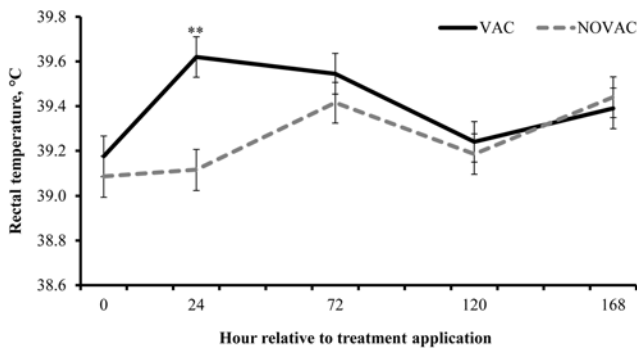


Figure 1. Rectal temperature of *Bos indicus* beef cows vaccinated (VAC; $n = 20$) or not vaccinated (NOVAC; $n = 20$) against the foot-and-mouth disease virus (5 mL subcutaneous of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, Brazil). A treatment \times hour interaction was detected ($P < 0.01$). **Treatment comparison within hour, $P \leq 0.01$.

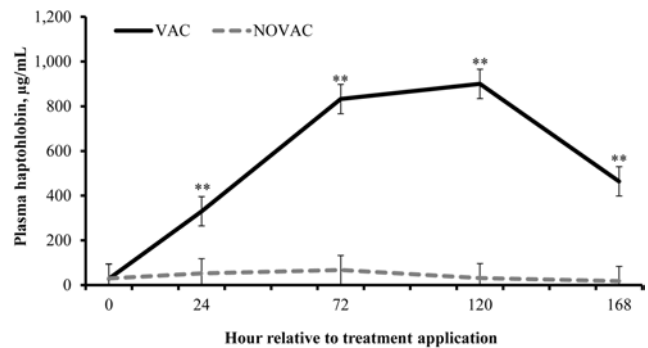


Figure 2. Plasma haptoglobin concentrations in *Bos indicus* beef cows vaccinated (VAC; $n = 20$) or not vaccinated (NOVAC; $n = 20$) against the foot-and-mouth disease virus (5 mL subcutaneous of Ourovac Aftosa; Ourofino Saúde Animal, Cravinhos, Brazil). A treatment \times hour interaction was detected ($P < 0.01$). **Treatment comparison within hour, $P \leq 0.01$.

with NOVAC cows at 24 h after treatment administration, and similar ($P \geq 0.31$) between treatments at 72, 120, and 168 h after treatment administration (Fig. 1).

A treatment \times hour interaction was also detected ($P < 0.01$) for plasma haptoglobin concentration, which was similar ($P = 0.98$) between VAC and NOVAC cows before treatment administration ($P = 0.48$) and greater ($P < 0.01$) in VAC compared with NOVAC cows at 24, 72, 120, and 168 h after treatment administration (Fig. 2).

DISCUSSION

In FMD endemic regions, cattle are vaccinated against the FMD virus every 6 mo due to the vaccine immune protection length (Parida, 2009). On the basis of the productive cycle of cow-calf operations (Hixon and Sanson, 2012), vaccination against the FMD virus often occurs during or shortly after the annual breeding season. Although previous research reported that FMD vaccines impair cattle production traits, including decreased milk production (Yeruham et al., 2001) and increased carcass lesions (Leal et al., 2014), the impacts of vaccination against the FMD virus on reproductive performance of beef cows still warranted investigation. Therefore, results from Exp. 1 are novel and support our hypothesis that administering a FMD vaccine during early pregnancy increased the incidence of pregnancy loss in beef cows 4-fold when compared with vaccine administration before timed AI. It is important to mention that these outcomes were independent of cow BCS and should not be associated with cow nutritional status during the estrus synchronization plus timed AI protocol (Cooke et al., 2009).

The majority of FMD vaccines utilized worldwide contain inactivated FMD virus serotypes and an oil-based adjuvant that elicits innate immune responses associated with antigen presentation to T-cell lymphocytes, including inflammatory and acute-phase reactions (Tizard, 2004; Rodriguez and Grubman, 2009; Rodrigues

et al., 2015). These immune responses, however, have been negatively associated with pregnancy maintenance (Hansen et al., 2004) and overall reproductive performance in cattle (Cooke et al., 2009). More specifically, adjuvants stimulate synthesis of proinflammatory cytokines (Rodrigues et al., 2015), which in turn elicit 2 major acute-phase responses: 1) synthesis of prostaglandins that lead to hyperthermia and 2) altered liver metabolism and gene regulation, favoring hepatic synthesis of acute-phase proteins such as haptoglobin (Carroll and Forsberg, 2007). Proinflammatory cytokines are known to impact pregnancy maintenance via direct embryotoxic effects and reduced endometrial cell proliferation in addition to increased body temperature and endometrial $\text{PGF}_{2\alpha}$ synthesis to levels that interrupt early pregnancy (Hansen et al., 2004). Conversely, haptoglobin does not have detrimental effects on cattle productive and reproductive functions, although this acute-phase protein is widely used to monitor inflammatory and acute-phase responses in cattle (Horadagoda et al., 1999; Cooke and Arthington, 2013).

Supporting this rationale, results from Exp. 2 demonstrated that administering a FMD vaccine elicited inflammatory and acute-phase responses, represented by treatment effects on rectal temperature and plasma haptoglobin concentrations, which can be directly associated with treatment effects detected for pregnancy loss in Exp. 1. Supporting findings from Exp. 2, Arthington et al. (2013) and Rodrigues et al. (2015) also reported that administering a vaccine containing inactivated pathogens plus adjuvant to beef cattle increased plasma haptoglobin concentrations for up to 120 h and associated these outcomes with reduced performance traits. It is important to note that inflammatory and acute-phase responses may also impair cattle fertility parameters such as follicle development and ovulation (Peter et al., 1989; Battaglia et al., 2000; Williams et al., 2001), which were not directly assessed in the present experiment, although pregnancy rates to AI on d 30

were similar between treatments. Given that the FMD vaccine utilized herein increased plasma haptoglobin concentrations for at least 7 d, it seems plausible that beef cows should be vaccinated against the FMD virus at least 1 wk before the beginning of the breeding season to prevent fertility and pregnancy losses, which supports the practical application of the VACPRES treatment evaluated in Exp. 1. Nevertheless, research is still warranted to determine the most appropriate timing for FMD vaccination to beef females.

In summary, administering a FMD vaccine to Nelore beef cows resulted in a 4-fold increase in pregnancy loss when vaccination occurred 30 d after timed AI compared with 31 d before timed AI. These outcomes can be associated with the inflammatory and acute-phase reactions elicited by the FMD vaccine, which are known to impair pregnancy maintenance in cattle (Hansen et al., 2004). Therefore, beef cows should not receive FMD vaccines containing inactivated virus and an oil-based adjuvant during early gestation; these vaccines should be administered before the beginning of the breeding season to prevent early pregnancy losses and optimize reproductive and overall efficiency of cow-calf operations.

REFERENCES

- Animal and Plant Health Inspection Service. 2013. Foot-and-mouth disease. http://www.aphis.usda.gov/publications/animal_health/2013/fs_fmd_general.pdf. (Accessed 7 July 2015.)
- Arthington, J. D., R. F. Cooke, T. D. Maddock, D. B. Araujo, P. Moriel, N. DiLorenzo, and G. C. Lamb. 2013. Effects of vaccination on the acute-phase protein response and measures of performance in growing beef calves. *J. Anim. Sci.* 91:1831–1837. doi:10.2527/jas.2012-5724.
- Battaglia, D. F., H. B. Krasa, V. Padmanabhan, C. Viguie, and F. J. Karsch. 2000. Endocrine alterations that underlie endotoxin-induced disruption of the follicular phase in ewes. *Biol. Reprod.* 62:45–53. doi:10.1095/biolreprod62.1.45.
- Brown, F. 1992. New approaches to vaccination against foot-and-mouth disease. *Vaccine* 10:1022–1026. doi:10.1016/0264-410X(92)90111-V.
- Carroll, J. A., and N. E. Forsberg. 2007. Influence of stress and nutrition on cattle immunity. *Vet. Clin. North Am. Food. Anim. Pract.* 23:105–149.
- Cooke, R. F., and J. D. Arthington. 2013. Concentrations of haptoglobin in bovine plasma determined by ELISA or a colorimetric method based on peroxidase activity. *J. Anim. Physiol. Anim. Nutr.* 97:531–536. doi:10.1111/j.1439-0396.2012.01298.x.
- Cooke, R. F., J. D. Arthington, D. B. Araujo, and G. C. Lamb. 2009. Effects of acclimation to human interaction on performance, temperament, physiological responses, and pregnancy rates of Brahman-crossbred cows. *J. Anim. Sci.* 87:4125–4132. doi:10.2527/jas.2009-2021.
- Curran, S., R. A. Pierson, and O. J. Ginther. 1986. Ultrasonographic appearance of the bovine conceptus from days 20 through 60. *J. Am. Vet. Med. Assoc.* 189:1295–1302.
- Federation of Animal Science Societies. 2010. Guide for the care and use of agricultural animals in agricultural research and teaching. 3rd ed. Fed. Anim. Sci. Soc., Savoy, IL.
- Grubman, M. J., and B. Baxt. 2004. Foot-and-mouth disease. *Clin. Microbiol. Rev.* 17:465–493. doi:10.1128/CMR.17.2.465-493.2004.
- Hansen, P. J., P. Soto, and R. P. Natzke. 2004. Mastitis and fertility in cattle—Possible involvement of inflammation or immune activation in embryonic mortality. *Am. J. Reprod. Immunol.* 51:294–301. doi:10.1111/j.1600-0897.2004.00160.x.
- Hixon, D. L., and D. W. Sanson. 2012. The Biological Cycle of the Beef Cow. In: *Cattle producer's handbook*. 3rd ed. J. R. Adams Publishing, Boise, ID. p. 400.
- Horadagoda, N. U., K. M. Knox, H. A. Gibbs, S. W. Reid, A. Horadagoda, S. E. Edwards, and P. D. Eckersall. 1999. Acute phase proteins in cattle: Discrimination between acute and chronic inflammation. *Vet. Rec.* 144:437–441. doi:10.1136/vr.144.16.437.
- Humbolt, P. 2001. Use of pregnancy specific proteins and progesterone assays to monitor pregnancy and determine the timing, frequencies and sources of embryonic mortality in ruminants. *Theriogenology* 56:1417–1433. doi:10.1016/S0093-691X(01)00644-6.
- Kahn, S., D. W. Geale, P. R. Kitching, A. Bouffard, D. G. Allard, and J. R. Duncan. 2002. Vaccination against foot-and-mouth disease: The implications for Canada. *Can. Vet. J.* 43:349–354.
- Leal, P. V., R. C. Pupin, A. C. Santos, T. C. Faccin, E. Surdi, C. R. B. Leal, R. C. Brumatti, and R. A. A. Lemos. 2014. Estimates of economic losses caused by local granulomatous reaction after use of an oily vaccine against FMD in cattle of Mato Grosso do Sul. *Pesqui. Vet. Bras.* 34:738–742. doi:10.1590/S0100-736X2014000800005.
- Leforban, Y. 1999. Prevention measures against foot-and-mouth disease in Europe in recent years. *Vaccine* 17:1755–1759. doi:10.1016/S0264-410X(98)00445-9.
- Meneghetti, M., O. G. Sa Filho, R. F. G. Peres, G. C. Lamb, and J. L. M. Vasconcelos. 2009. Fixed-time artificial insemination with estradiol and progesterone for *Bos indicus* cattle: I. Basis for development of protocols. *Theriogenology* 72:179–189. doi:10.1016/j.theriogenology.2009.02.010.
- Parida, S. 2009. Vaccination against foot-and-mouth disease virus: Strategies and effectiveness. *Expert Rev. Vaccines* 8:347–365. doi:10.1586/14760584.8.3.347.
- Peter, A. T., W. T. K. Bosu, and R. J. DeDecher. 1989. Suppression of preovulatory luteinizing hormone surges in heifers after intrauterine infusions of *Escherichia coli* endotoxin. *Am. J. Vet. Res.* 50:368–373.
- Rodrigues, M. C., R. F. Cooke, R. S. Marques, B. I. Cappelozza, S. A. Arispe, D. H. Keisler, and D. W. Bohnert. 2015. Effects of vaccination against respiratory pathogens on feed intake, metabolic, and inflammatory responses in beef heifers. *J. Anim. Sci.* 93:4443–4452. doi:10.2527/jas.2015-9277.
- Rodriguez, L. L., and C. G. Gay. 2011. Development of vaccines toward the global control and eradication of foot-and-mouth disease. *Expert Rev. Vaccines* 10:377–387. doi:10.1586/erv.11.4.
- Rodriguez, L. L., and M. J. Grubman. 2009. Foot and mouth disease virus vaccines. *Vaccine* 27:D90–D94. doi:10.1016/j.vaccine.2009.08.039.
- Tizard, I. R. 2004. Vaccines and their production. In: T. Merchant, editor, *Veterinary immunology*. 7th ed. Elsevier, Philadelphia, PA. p. 247.
- Wagner, J. J., K. S. Lusby, J. W. Oltjen, J. Rakestraw, R. P. Wettemann, and L. E. Walters. 1988. Carcass composition in mature Hereford cows: Estimation and effect on daily metabolizable energy requirement during winter. *J. Anim. Sci.* 66:603–612.
- Williams, C. Y., T. G. Harris, D. F. Battaglia, C. Viguie, and F. J. Karsch. 2001. Endotoxin inhibits pituitary responsiveness to gonadotropin-releasing hormone. *Endocrinology* 142:1915–1922.
- Yeruham, I., H. Yadin, M. Haymovich, and S. Perl. 2001. Adverse reactions to FMD vaccine. *Vet. Dermatol.* 12:197–201. doi:10.1046/j.0959-4493.2001.00221.x.